

# Feature annotation

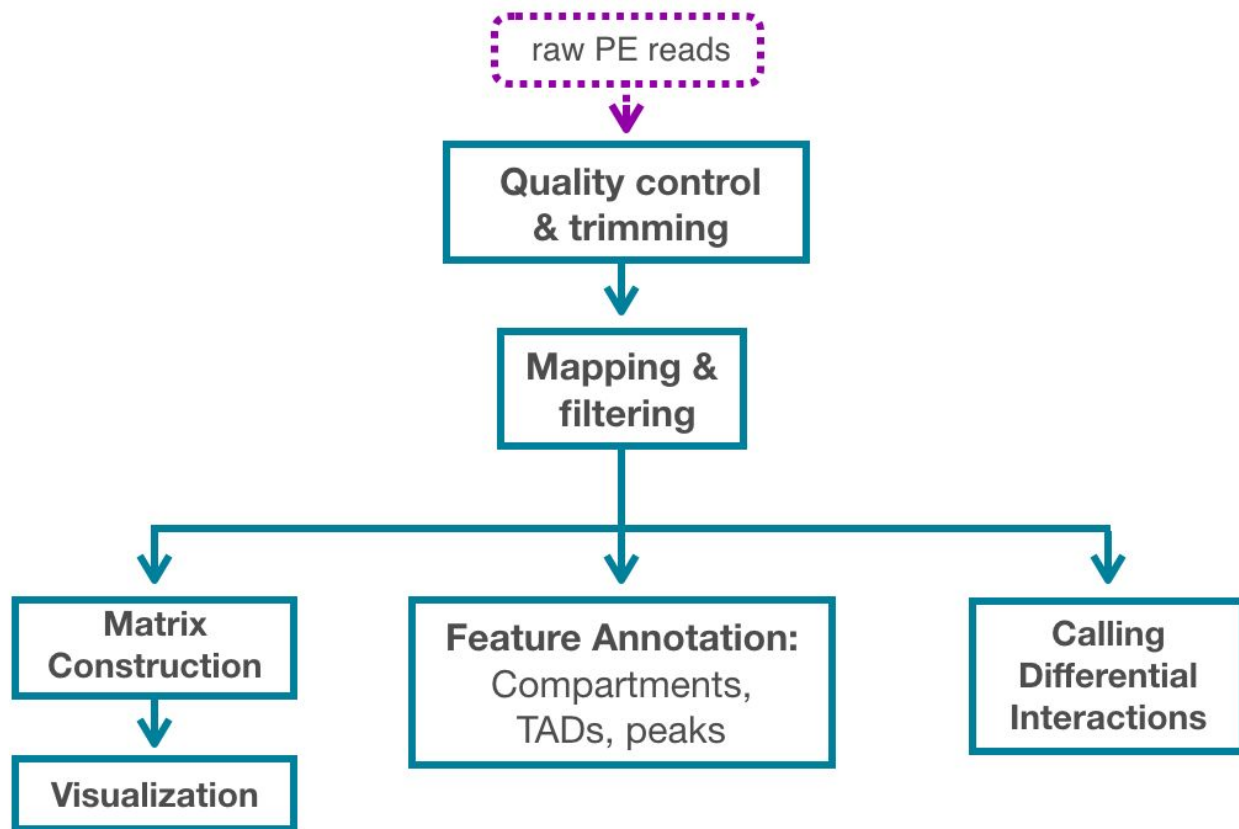
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Compartments, TADs and peaks

# Compartments

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# Recap

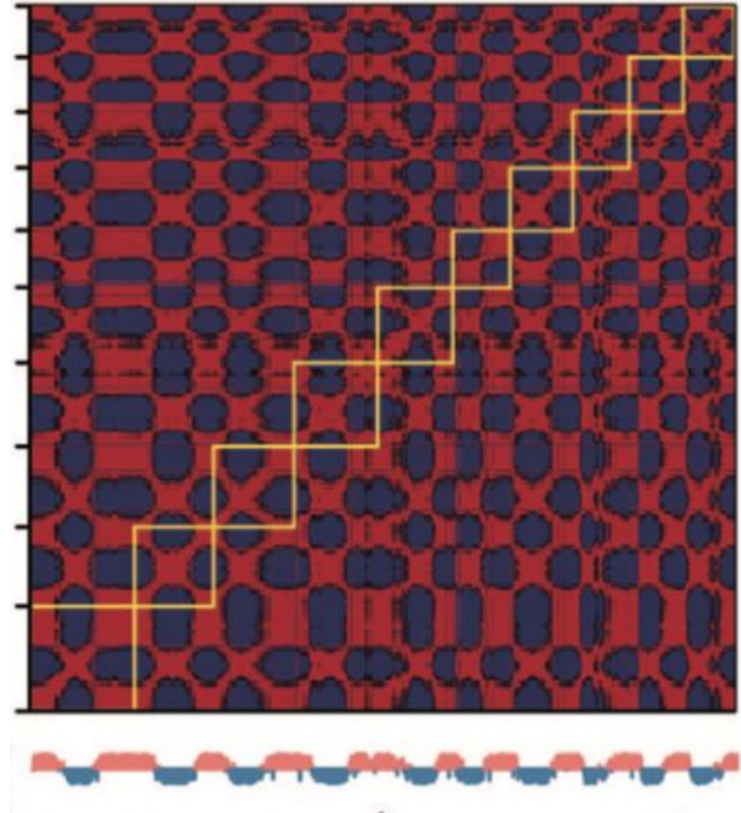


# Learning objectives

- Identify compartments in HiC matrix
- Compartments in plants

# Compartments

- A compartment:
  - active regions (euchromatin)
- B compartment:
  - inactive regions (heterochromatin)
- Bin sizes 100kb - 1MB



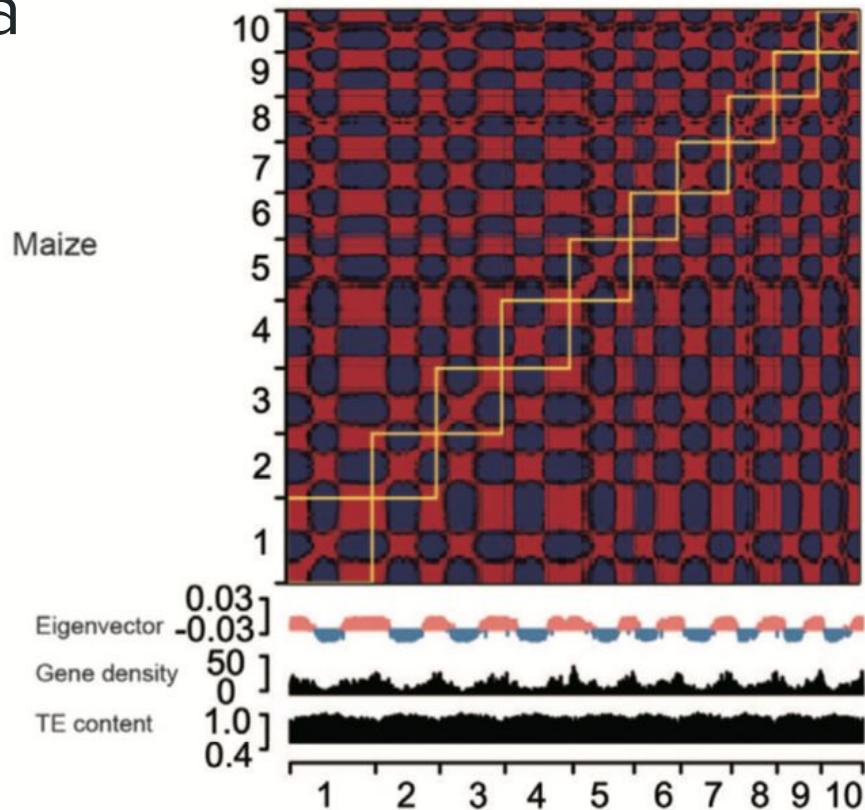
- PC1

# PCA

- Principal Component Analysis
- The first eigenvector will give the compartmentalization profile
- Positive values indicate one compartment, negative values indicate other compartment
- How to tell if A or B?

# Correlation with other data

- Show first component together with other epigenomic tracks:
- Gene content / TE density
- Histone modifications:
  - A: H3K27Ac, H3K4me3
  - B: H3K27me3, H3K9me3
- Transcription
  - RNA seq tracks



# Practical

- Identify compartments with HiCexplorer

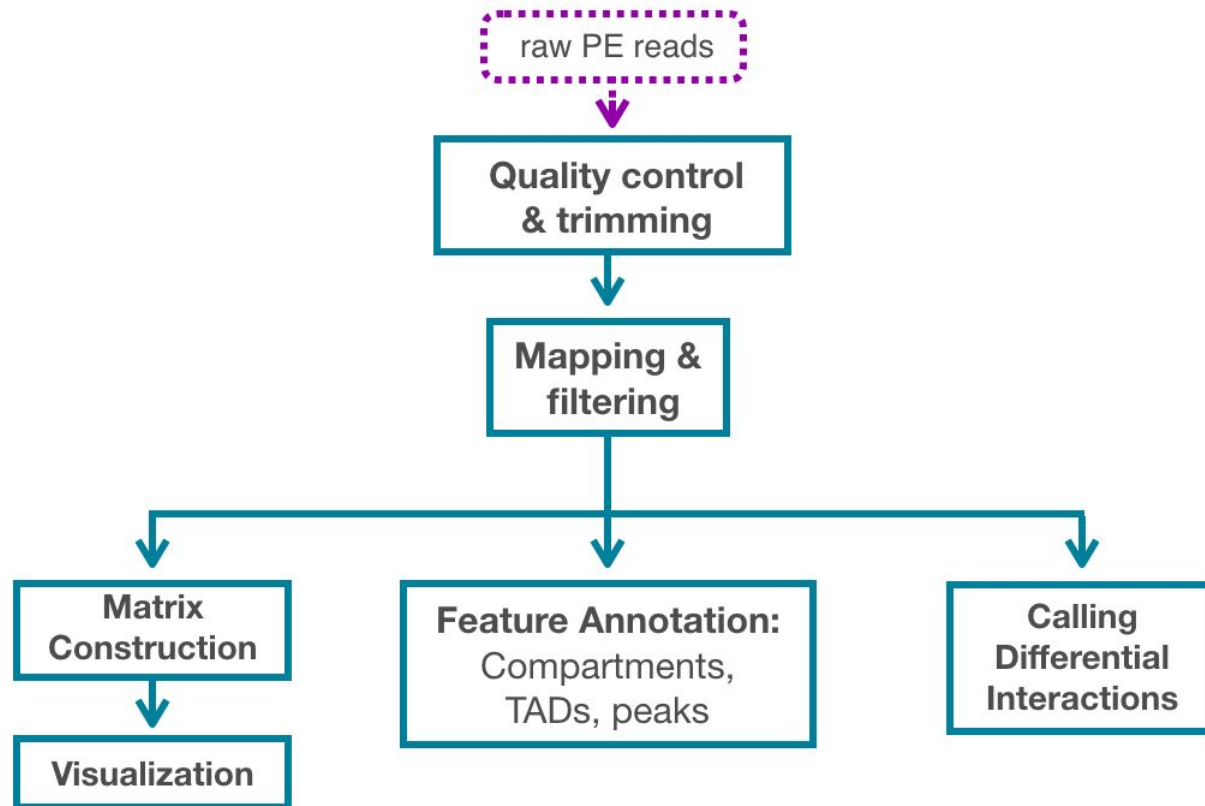




TADs

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# Recap

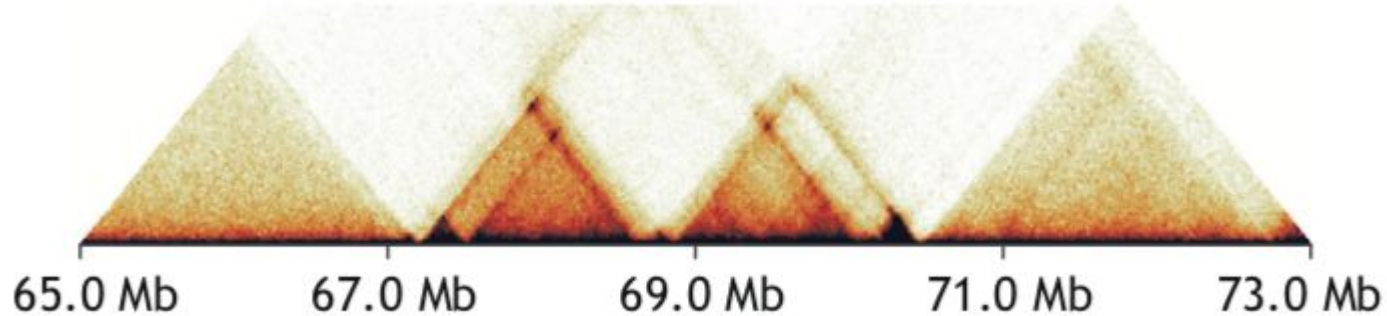


# Learning objectives

- Identify TADs with HiCExplorer
- Discuss challenges in TAD calling

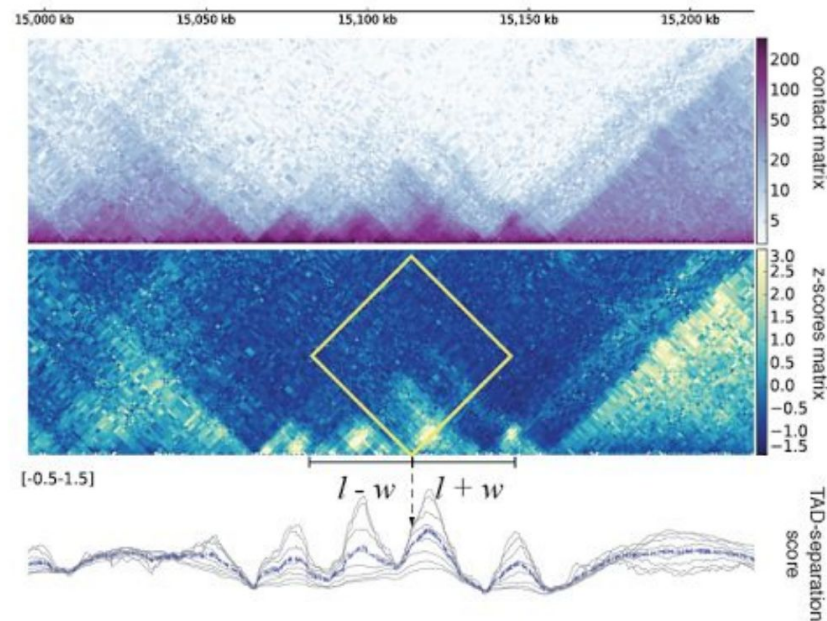
# TADs

- TADs are regions with elevated self interaction frequencies
- TADs might act as an insulated genomic region that constrains regulatory interactions



# TAD calling in HiCexplorer

1. Transform matrix to Z score (subtract mean contact frequency at each distance): make bins more comparable
2. For each bin, calculate average contacts between  $w$  upstream and downstream bins.
3. Repeat for different values of  $w$ , and then take the average.
4. Local minima should be boundaries!
5. To double check: compare the distributions of upstream and downstream “diamonds”.



# TAD calling challenges

- What is a TAD?
- TAD-like patterns are often hierarchical and overlapping (subTADs, gene mini domains)
- There are aprox. 22 different TAD calling algorithms
  - Current approaches often not reproducible
  - Sensible to normalization, bin size, sequencing depth

RESEARCH

Open Access

## Comparison of computational methods for the identification of topologically associating domains



Marie Zufferey<sup>1,2†</sup>, Daniele Tavernari<sup>1,2†</sup>, Elisa Oricchio<sup>3</sup> and Giovanni Ciriello<sup>1,2\*</sup> 

# TAD calling algorithms

- Linear score
  - Directionality Index
  - Insulation score
    - TopDom
    - **HiCEXplorer**
- Statistical models
  - TADbit
- Clustering
  - ClusterTAD
- Network analysis
  - spectral

# Practical:

- TAD calling with HiCexplorer

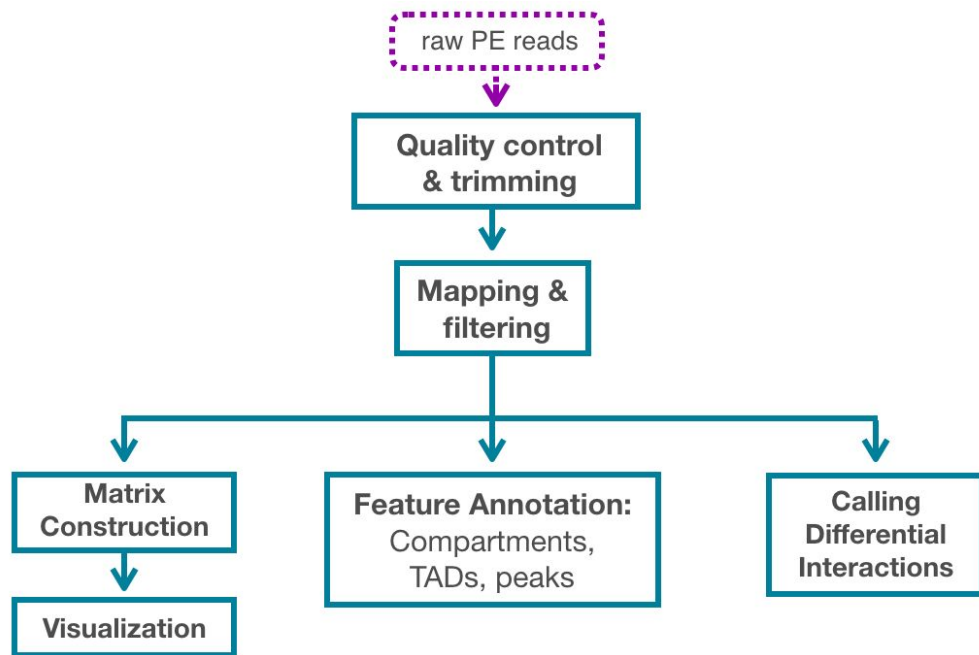




# Interaction peaks

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# Recap

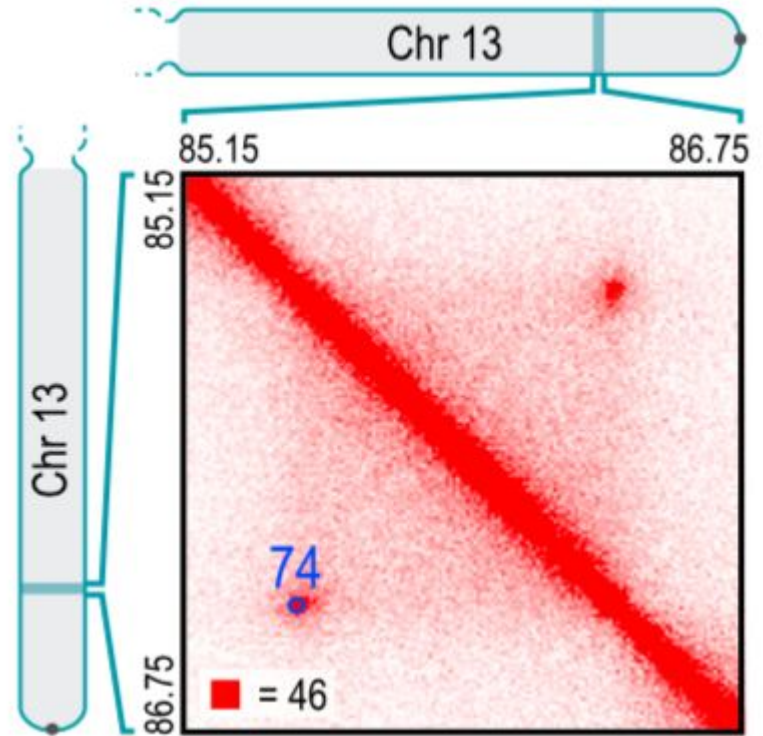


# Learning objectives

- Identify peaks with HiCCUPS

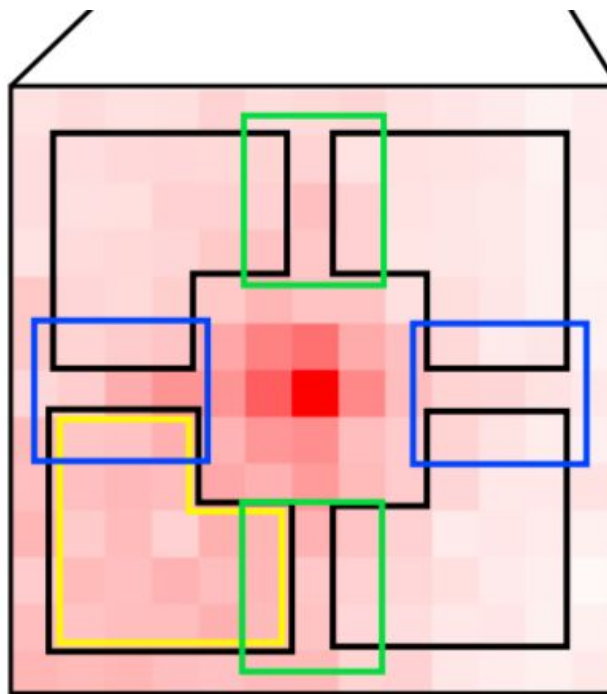
# Peaks and biological features

- Enhancer-promoter interactions
- CTCF-CTCF and cohesin binding sites
- Polycomb bodies
- KNOT region (Arabidopsis)
- Gene - Gene interactions (Maize)



# HICCUPS

- Local enrichment over four backgrounds:
  - Donut
  - Horizontal
  - Vertical
  - Lower left corner
- Compare bin contact frequency with average contact frequency of each background



$$\text{Exp}(\text{yellow box}) = 83.3$$

$$\text{Exp}(\text{black box}) = 67.4$$

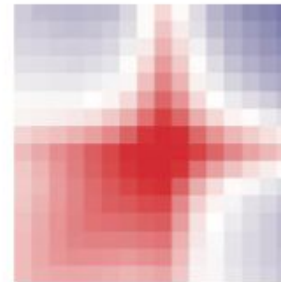
$$\text{Exp}(\text{blue box}) = 78.9$$

$$\text{Exp}(\text{green box}) = 88.4$$

$$\text{Peak} = 341$$

# Aggregate Peak Analysis

- Analyse the average interaction profile for all peaks
- Can be used as QC



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Loops

# Peak calling algorithms

- Global enrichment
  - Fit-HiC
  - HOMER
  - HICCUPS
- Local enrichment
  - **HICCUPS**
  - **HiCExplorer**

# Practical:

- Peak calling with HiCexplorer





# Resources:

- HiCExplorer: <https://hicexplorer.readthedocs.io/>
- HiGlass: <http://higlass.io/>
- Deeptools: <https://deeptools.readthedocs.io/en/develop/>
- Juicer: <https://github.com/aidenlab/juicer/wiki>
- Collection of hic tools: [https://github.com/mdozmorov/HiC\\_tools](https://github.com/mdozmorov/HiC_tools)